



# APOE signaling in neurodegenerative diseases: an integrative approach targeting APOE coding and noncoding variants for disease intervention

Xiaopu Zhou<sup>1,2,3</sup>, Amy KY Fu<sup>1,2,3</sup> and Nancy Y Ip<sup>1,2,3</sup>

*APOE (apolipoprotein E)* is a key regulator of lipid metabolism and a leading genetic risk factor for Alzheimer's disease. While *APOE* participates in multiple biological pathways, its roles in diseases are largely due to the mutant protein encoded by *APOE-ε4*. However, emerging evidence suggests that some noncoding Alzheimer's disease risk variants residing in *APOE* and its nearby regions exert *APOE-ε4*-independent risks and modify *APOE* gene expression. Moreover, intervention strategies targeting *APOE* are being explored. In this review, we summarize the literature on the genetic risks and roles of *APOE* in biological systems. Moreover, we propose an integrative approach to evaluate disease risk and tailor interventions to aid research on *APOE*-associated diseases.

## Addresses

<sup>1</sup> Division of Life Science, State Key Laboratory of Molecular Neuroscience, Molecular Neuroscience Center, The Hong Kong University of Science and Technology, Hong Kong, China

<sup>2</sup> Hong Kong Center for Neurodegenerative Diseases, Hong Kong Science Park, Hong Kong, China

<sup>3</sup> Guangdong Provincial Key Laboratory of Brain Science, Disease and Drug Development, Hong Kong University of Science and Technology Shenzhen Research Institute, Shenzhen-Hong Kong Institute of Brain Science, 518057 Shenzhen, Guangdong, China

Corresponding author: Ip, Nancy Y ([boip@ust.hk](mailto:boip@ust.hk))

**Current Opinion in Neurobiology** 2021, 69:58–67

This review comes from a themed issue on **Molecular neuroscience**

Edited by **Frank Bradke** and **Yukiko Goda**

<https://doi.org/10.1016/j.conb.2021.02.001>

0959-4388/© 2021 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

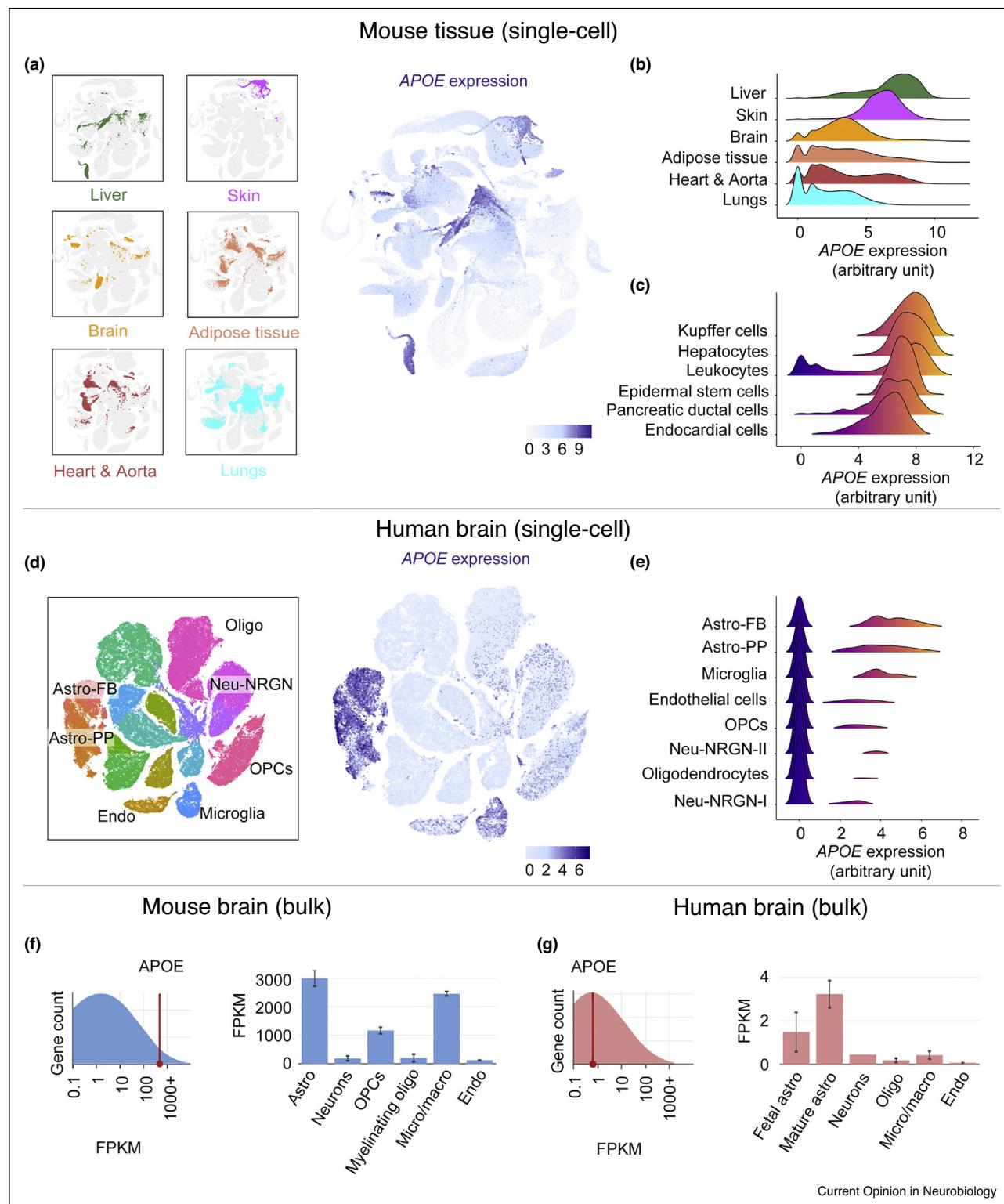
## Introduction

ApoE (apolipoprotein E), encoded by *APOE*, was first described as an arginine-rich polypeptide [1–4]. ApoE is a key regulator of triglyceride and cholesterol metabolism in the peripheral system and brain [5]. In the peripheral system, plasma ApoE mainly resides in lipoprotein particles including very-low-density lipoproteins, chylomicrons, and subsets of high-density lipoproteins,

facilitating the hepatic endocytosis of intestinal chylomicrons and the lipid redistribution across tissues [6–8]. ApoE is also highly expressed in the brain, which contains the largest proportion of cholesterol in the body (~20% of the total) [9]. In the brain, ApoE mainly exists in high-density, lipoprotein-like particles and maintains lipid homeostasis and proper neuronal functioning [10]. Human and mice lacking *APOE* exhibit functional deficits in both the peripheral system and brain. In particular, *APOE*-deficient mice exhibit elevated plasma cholesterol levels, early onset of atherosclerotic lesions, and elevated inflammation [11]. Moreover, compared to wild-type mice, *APOE*-deficient mice exhibit fewer dendrites and lower synapse density in neurons of the neocortex and hippocampus as well as greater age-dependent neurodegeneration in the neocortex [12]. Meanwhile, humans lacking *APOE* exhibit exceptionally high cholesterol levels; mild atherosclerosis; normal cognitive function with subtle deficits in memory, language, visuospatial abilities and executive functions [13•]. Hence, ApoE has important roles in maintaining normal physiological functions in both the brain and peripheral system.

## ***APOE*-associated pathways in the brain and peripheral system**

ApoE affects diverse biological pathways in both the nervous and immune systems that are associated with neurodegenerative diseases. Biochemical analysis suggests that *APOE* is predominantly expressed by the liver, adipose tissue, arteries, and brain [6]. Single-cell RNA sequencing technique enable high-throughput assays of comprehensive gene expression profiles at single-cell resolution. Accordingly, reanalysis of publicly available data with mouse tissues revealed that *APOE* is predominantly expressed in the liver, skin, brain, adipose tissue, heart, aorta, and lungs, particularly in Kupffer cells, hepatocytes, and leukocytes [14]. Kupffer cells (a type of resident hepatic macrophage) along with hepatocytes are the major sources of peripheral *APOE* expression, although the role of *APOE* expressed by Kupffer cells is not well understood [15]. Meanwhile, in the human brain, *APOE* is mainly expressed by astrocytes, microglia, oligodendrocyte progenitors and oligodendrocytes, endothelial cells, and NRGN-expressing neurons (Figure 1) [16]. *NRGN* encodes neurogranin, a synaptic protein associated with synaptic strength whose level in cerebrospinal fluid is altered in patients with neurodegeneration or Alzheimer's disease (AD) [17–19]. Moreover, according

**Figure 1**

APOE expression profiles at the single-cell and tissue levels.

(a–c) APOE expression in mouse tissues and cells. APOE expression profiles in selected mouse tissues at the (a) single-cell level and (b) tissue level. (c) APOE expression profiles in selected cell types. (d, e) APOE expression in human brain cells. (d) APOE expression profiles at the single-cell level in the human brain prefrontal cortex and anterior cingulate cortex. (e) APOE expression profiles in selected cell types. (f, g) APOE expression in (f) the mouse and (g) human brain. Histogram showing the ranking of the APOE transcript levels across all detected genes in the

Current Opinion in Neurobiology

to the Brain RNA-Seq database (<https://www.brainrnaseq.org/>) that integrates bulk RNA-Seq data from mouse and human brains [20,21], *APOE* expression is higher in the mouse brain than the human brain (Figure 1). These lines of evidence corroborate the findings of biochemical studies on *APOE* expression profiles, providing a basis for further characterization of the roles of *APOE* and associated pathways in neurodegenerative diseases.

Most studies on *APOE* in the brain are restricted to neurons, astrocytes, and microglia, which have revealed specific function of *APOE* signaling in those cells (Table 1). Astrocytes are the major source of the *APOE* in the brain, which can modulate neurite outgrowth [22]. The activation of *APOE* signaling in neurons triggers phosphorylation cascades that activate several downstream kinases [23]. Specifically, *APOE* signaling modulates neurite outgrowth/extension, synaptogenesis, axon remodeling [24–26], and calcium homeostasis [27] and also prevents neuronal apoptosis [28]. Furthermore, as a protective mechanism induced upon injury or stress, neurons increase their *APOE* expression to recruit lipids from the surrounding environment for repair [8]. Moreover, upon activation, microglia exhibit elevated *APOE* expression [29], which modulates their molecular phenotypes and regulates their cell state transition from a homeostatic state to a disease/neurodegeneration-associated state by influencing gene expression in a *TREM2*-dependent manner [30•]. Furthermore, ApoE4, a mutated form of ApoE protein that is known to associate with AD pathogenesis, can bind to DNA and act as a transcription factor to modulate cytokine production in neuronal cell lines, further indicating that *APOE* plays important roles in transcriptomic regulation of immune-related pathways [31,32]. Notably, a recent human postmortem brain study revealed the presence of *APOE* circular RNA, which accounts for approximately one-third of the total brain *APOE* RNA, although its function and cellular expression profile are unknown [33•]. Therefore, further investigation is required to understand the roles of *APOE* expression and its associated pathways in the brain.

Emerging studies also suggest that *APOE* plays roles in the vascular system. The breakdown of blood–brain barrier (BBB) might account for the neurodegeneration and neuroinflammation in AD [34]; the *APOE*-deficient mice exhibit age-dependent BBB disruption [35]. In addition, disease-associated ApoE4 protein can also lead to BBB dysfunction [36,37]. Moreover, a recent study using an induced pluripotent stem cell-derived BBB

model suggests that ApoE4 expressed by pericytes might modulate amyloid deposition near the BBB, making it a possible trigger of microvasculature injury [38•]. Interestingly, although peripheral ApoE cannot effectively diffuse into the brain owing to its limited BBB permeability [39–41], manipulating peripheral ApoE level can potentially affect cognitive functions [42•]. Given the modulatory effects of both vascular and peripheral *APOE* on brain functions, an intervention strategy targeting these sources of *APOE* could be effective for certain neurodegenerative diseases including AD.

### ***APOE* coding variants in human diseases**

Genetics and biochemical studies revealed the existence of 3 *APOE* isoforms in the general population—*APOE*-ε2 (encoding ApoE2; Cys112–Cys158), *APOE*-ε3 (the most common form; Cys112–Arg158), and *APOE*-ε4 (encoding ApoE4; Arg112–Arg158)—which are defined by combinations of 2 coding mutations in exon 4 of *APOE* [43–47] (Table 2). These mutations of *APOE* modify its protein function by altering internal domain–domain interactions [48]. In ApoE4, the cysteine-to-arginine substitution at residue 112 renders the formation of a new salt bridge between Glu109 and Arg112, and modifies the orientation of Arg61 (a key residue), resulting in ApoE4 having higher affinity for very-low-density lipoproteins [49,50]. Meanwhile, in ApoE2, the arginine-to-cysteine substitution at residue 158 changes the salt bridge within and between the helical structures, which also modifies the charges of the receptor-binding region [50]. Accordingly, ApoE4 does not efficiently interact with high-density lipoproteins and preferentially binds low-density lipoproteins, whereas ApoE2 exhibits greatly reduced affinity for low-density lipoprotein receptor [49,51].

An early genetics study revealed an association between *APOE* and abnormal lipid metabolism [8]. Specifically, *APOE*-ε2 is one of the genetic risk factors of type III hyperlipoproteinemia, a familial lipoprotein metabolic disorder characterized by elevated blood cholesterol and triglyceride levels along that incurs risks of severe atherosclerotic vascular diseases [52]. In contrast, *APOE*-ε2 is also associated with longevity, reduced age-associated cognitive decline, and reduced AD risk [53]. Meanwhile, ApoE-ε4 is associated with neurodegenerative diseases including AD, vascular dementia, Lewy body dementia, frontotemporal dementia, Parkinson’s disease, and multiple sclerosis; some cerebrovascular disorders such as cerebral amyloid angiopathy and stroke; and poor outcomes after traumatic brain injury [54]. Given that these diseases are predominately associated

(Figure 1 Legend Continued) corresponding tissues. Data obtained from the Brain RNA-Seq database. (a, d right panels) *APOE* expression in individual cells was visualized by *t*-distributed stochastic neighbor embedding (*t*-SNE) plots; color denotes normalized *APOE* expression in individual cells. Astro, astrocytes; Astro-FB, fibrous astrocytes; Astro-PP, protoplasmic astrocytes; Endo, Endothelial cells; FPKM, fragments per kilobase of transcript per million mapped reads; Neu, neurons; Neu-NRGN, NRGN-expressing neurons; Oligo, oligodendrocytes; OPCs, oligodendrocyte progenitor cells; Macro, macrophages; Micro, microglia.

**Table 1****Roles of APOE-associated pathways in the brain and peripheral system**

Tissue or cell type	Function	References
Peripheral system	<ul style="list-style-type: none"> <li>• Redistribute lipids</li> <li>• Modify cognitive function</li> </ul>	[6–8] [42*]
Astrocytes	<ul style="list-style-type: none"> <li>• Produce majority of the ApoE protein in the brain</li> <li>• Modulate neurite outgrowth</li> </ul>	[8] [22]
Neurons	<ul style="list-style-type: none"> <li>• Modulate neurite outgrowth and extension, synaptogenesis, and axon remodeling</li> <li>• Regulate calcium homeostasis</li> <li>• Confer neuroprotection</li> <li>• Repair damaged neurons by recruiting lipids from the surrounding environment</li> <li>• Induce microglial activation</li> <li>• Regulate transcriptome profile</li> </ul>	[24–26] [27] [28] [8] [30*]
Microglia		

**Table 2****Summary of APOE isoforms and associated phenotypes**

Isoform (or alias)	Mutations	Phenotypes/Notes	References
$\epsilon 1$ (Weisgraber allele)		Obesity Non insulin-dependent diabetes Hypertension Moderate lipid disturbances	[64]
$\epsilon 1y$ ( $\epsilon 3r$ )	Asp127–Cys158	No obvious phenotypic change	[65]
$\epsilon 2$	Arg112–Cys158	Longer lifespan	
	Cys112–Cys158	Type III hyperlipoproteinemia (elevated blood cholesterol and triglyceride levels) Atherosclerotic vascular diseases Reduced risk of Alzheimer's disease	[52,53]
$\epsilon 3$	Cys112–Arg158	Most frequent isoform in the general population	
$\epsilon 4$	Arg112–Arg158	Neurodegenerative diseases including Alzheimer's disease Cerebrovascular disorders	[54]
$\epsilon 5$	Lys3	Mild hypertriglyceridemia	[66]
$\epsilon 5$	Arg84–Arg112	Elevated blood cholesterol level	
$\epsilon 7$	Lys244–Lys245	Triple-vessel disease Diabetes mellitus Hypertriglyceridemia	[67]
Christchurch allele (R136S)	Ser136	Significant enrichment of mutated ApoE in very low-density lipoprotein Type III hyperlipoproteinemia	[68,72**]

with the nervous and vascular systems and involve altered immune status, *APOE* might have broad roles in modulating specific pathways associated with neuronal, vascular, and immune functions.

Of note, *APOE*- $\epsilon 4$  is a well-recognized genetic risk factor for AD, which is one of the most common neurodegenerative diseases [55]. *APOE*- $\epsilon 4$  modulates various AD-associated endophenotypes covering a broad spectrum of disease signatures including synaptic plasticity [56], neuronal activity [57], brain volume [58], cognitive performance [59], onset age [60], and brain immune status [61]. Compared to *APOE*- $\epsilon 2$  and *APOE*- $\epsilon 3$ , *APOE*- $\epsilon 4$  can also modify the pathological hallmarks of AD, including amyloid-beta (A $\beta$ ), tau neurofibrillary tangles, and neurodegeneration by promoting A $\beta$  aggregation, inhibiting A $\beta$  clearance, and exacerbating tau-mediated neurodegeneration in tau transgenic mice [62,63]. Hence, *APOE*- $\epsilon 4$  plays critical roles in the

pathogenesis of AD by modulating a plethora of biological pathways involved in the disease pathogenesis.

Multiple rare *APOE* coding risk variants have also been identified, including the *APOE*- $\epsilon 1$  Weisgraber allele (Asp127–Cys158) [64], *APOE*- $\epsilon 1y$  (Arg112–Cys158) [65], several forms of *APOE*- $\epsilon 5$  (including Lys3 and Arg84–Arg112) [66,67], *APOE*- $\epsilon 7$  (Lys244–Lys245) [68], and the Christchurch mutation (R136S or Ser136) [69], which potentially modify disease risk by altering ApoE function (Table 2). For instance, ApoE with the Christchurch mutation tends to exist in a higher-order oligomerization state with a larger aggregate size when compared with the normal ApoE protein [70], which consequently reduces the binding affinity of *APOE* for low-density lipoprotein receptor by 60% [71]. In particular, in a case report of a homozygous carrier who exhibited high brain A $\beta$  load, the Christchurch mutation elicited a strong neuronal protective effect against the causal *PSEN1*

E280A coding risk variant, as indicated by low tau tangle load, low plasma neurofilament light chain level, and mild cognitive issues [72<sup>••</sup>]. This implies that appropriately manipulating the *APOE* signaling pathway alone might be sufficient for AD intervention.

Notably, besides altering protein function, mutations of *APOE*-ε2 and *APOE*-ε4 are located in a well-defined genomic region with DNA methylation modification (i.e., a CpG island) [73]. Moreover, an *APOE*-ε4 variant introduces a cytosine base that is highly methylated (65–90%) across human tissues [74]. Concordantly, the *APOE*-ε4 allele is associated with reduced brain *APOE* expression [75,76<sup>••</sup>]. In addition, compared to noncarriers, *APOE*-ε4 heterozygous carriers exhibit decreased *APOE*-ε4 mRNA expression in the brain and blood [76<sup>••</sup>,77]. Therefore, *APOE*-ε4 potentially exerts disease risk effects by encoding a mutated protein with altered *APOE* expression and functions, which leads to the dysregulation of *APOE*-associated signaling pathways and consequently modifies disease risk.

### Contributions of *APOE* noncoding risk variants to Alzheimer's disease

*APOE* coding risk variants unequivocally impact associated signaling pathways, while the dysregulation of *APOE* expression might also trigger or modulate AD onset and progression. The expression of *APOE* is controlled in a tissue-specific manner by multiple enhancers in nearby regions [74,78], which might also be modified by variants located in noncoding regions [79–83]. Association analyses have revealed numerous AD-associated noncoding variants and haplotypes residing in *APOE* and its nearby regions [84–86]. Interestingly, several studies also show that a specific haplotype structure with an ethnic-specific distribution might account for the variable AD risk of *APOE*-ε4 carriers among populations [87<sup>•</sup>,88<sup>•</sup>]. Nevertheless, it remains controversial whether those noncoding variants exert risk effects for AD.

Accordingly, our group conducted a comprehensive fine-mapping analysis of *APOE* genetic risk in *APOE* and its nearby regions based on whole-genome sequencing data [76<sup>••</sup>]. Our analysis enabled the identification of all genomic variants and their haplotype structures in the study cohort for association analysis, which identified noncoding risk variants and haplotypes residing in the *PVRL2* and *APOC1* regions that exert AD risk effects independent of *APOE*-ε4. Further analysis revealed their potential regulatory functions in the modification of brain *APOE* expression; specifically, carriers of *PVRL2* or *APOC1* haplotypes exhibited higher brain *APOE* expression than noncarriers. Moreover, if the noncoding risk variants and *APOE*-ε4 reside in the same chromosome, those noncoding variants might modify the disease penetrance of *APOE*-ε4 via enhanced expression of ApoE4. Meanwhile, the identified risk haplotypes are more

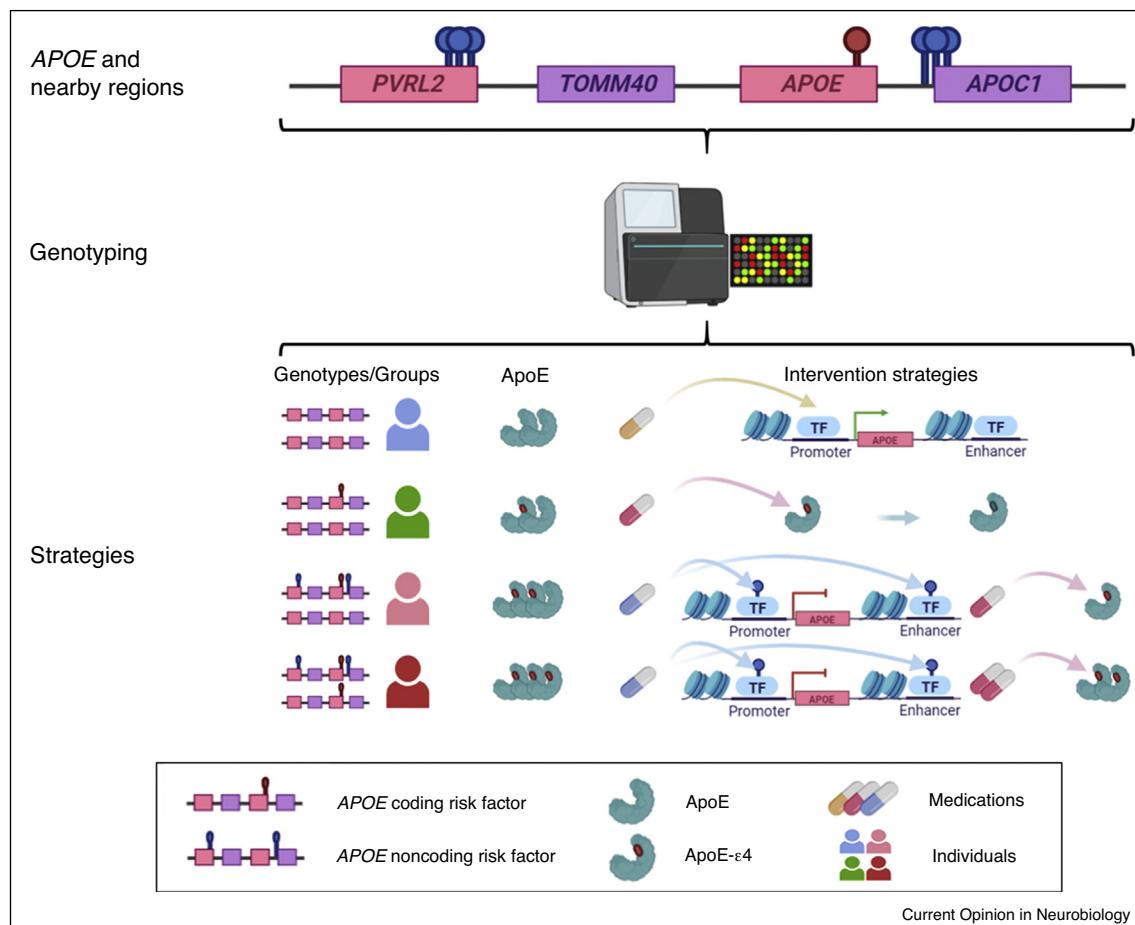
prevalent in populations of European descent than those of African descent, corroborating the lower AD risk of *APOE*-ε4 observed in the latter [89]. Taken together, the discoveries of these noncoding risk factors and their putative regulatory functions provide a genetic basis for the variable *APOE*-ε4 AD risk across ethnic groups and suggests that changes in *APOE* expression alone might also confer AD risk.

Notably, the AD-associated *APOE* noncoding variants span the *PVRL2*–*TOMM40*–*APOE*–*APOC1* region, potentially affecting the expressions of those genes and influencing several other AD-associated mechanisms in parallel to the Aβ cascade, including viral infection, mitochondrial dysfunction, and neuroinflammation [90–92]. Specifically, *PVRL2* encodes a surface protein that abets herpes and pseudorabies viruses entering cells—a process potentially associated with AD pathogenesis [93,94]. Meanwhile, *TOMM40* encodes a mitochondrial membrane protein that facilitates protein transport into mitochondria, potentially accounting for the observed mitochondrial dysfunction in the AD brain [95]. Furthermore, *APOC1* encodes a protein involved in lipid metabolism that is implicated in atherosclerosis, which is a possible trigger of neuroinflammation [96,97]. Hence, the identified noncoding *APOE* risk factors might elicit additional effects on neurodegenerative diseases in addition to modifying *APOE* expression.

### Proposed model for *APOE*-associated intervention strategies

Corroborating the critical involvement of *APOE* in AD pathogenesis, intervention strategies targeting *APOE* expression or ApoE4 mutant proteins can ameliorate AD-associated symptoms. For instance, administration of the RXR agonist bexarotene, which can trigger *APOE* expression, rapidly enhances Aβ clearance and improves neuronal and cognitive functioning in AD mouse models [98]. Meanwhile, PH002, a small molecule structure corrector that can reshape ApoE4 protein into an ApoE3-like structure, can ameliorate the neuronal toxicity of ApoE4 and reduce p-tau, Aβ<sub>40</sub>, and Aβ<sub>42</sub> levels in induced pluripotent stem cell-derived neurons [99<sup>•</sup>]. Moreover, other intervention strategies including ApoE-neutralizing antibody [100], *APOE* antisense oligonucleotides [101], and virus-mediated ApoE2 overexpression [102] can also elicit protective effects against AD by modulating ApoE2 expression and its accompanying signaling pathways. Thus, such *APOE*-targeting therapeutic strategies could be developed as treatments for neurodegenerative diseases.

Nevertheless, considering the complexity of *APOE* signaling, which involves multiple cell types, *APOE* signaling must be manipulated in a cell-type-specific and genotype-aware manner. Moreover, applying the aforementioned intervention strategies at a population scale might yield inconsistent effects owing to variations in

**Figure 2**

#### Integrative approaches targeting *APOE*-related diseases.

The coding and noncoding genotypes of *APOE* and its nearby regions are detected by array or sequencing methods. Individuals harboring different combinations of coding and noncoding mutations are segregated and subjected to different intervention paradigms using drugs targeting *APOE* expression and ApoE mutant proteins. TF, transcription factor.

genetic background. For instance, among AD patients treated with bexarotene, *APOE*-ε4 carriers exhibit no significant change in Aβ load, whereas *APOE*-ε3 homozygous patients exhibit a regional reduction in Aβ load [103•]. On the other hand, the presence of certain *APOE* noncoding genetic risk variants might modify the chromatin state of nearby regions and consequently interfere with the *APOE* expression-inducing effect of bexarotene [104]. Therefore, a rigorous investigation of the genomic contexts of *APOE* and nearby regions may facilitate the development of more-effective and genotype-specific intervention strategies. For instance, a multistage screening assay can be applied to individuals identified as having *APOE*-ε4, both *APOE*-ε4 and non-coding risk variants, or only noncoding risk variants—each of whom may be submitted to a different treatment paradigm for AD.

Accordingly, by combining coding and noncoding genetic information, a polygenic risk score model can be designed to stratify patients and assess an individual's risk of developing a given disease [105–107]. Furthermore, long-read sequencing technology can directly capture all genomic contexts of the *APOE* and its nearby regions, including single nucleotide polymorphisms, insertions, deletions, structural variations, and haplotype information, which would more accurately stratify individuals for subsequent preclinical or clinical investigation [76••]. The results of such research might yield tailored intervention strategies applicable to a broad range of diseases associated with *APOE* (Figure 2).

#### Conclusion

*APOE* is expressed in various cell types and involved in diverse biological functions. The contributions of *APOE* coding and noncoding genetic variants to human diseases

through the modulation of protein functions and gene expression highlight the roles of the dysregulation of *APOE* signaling in modifying disease risk. Meanwhile, *APOE*-targeting intervention strategies have demonstrated beneficial effects in decreasing disease-associated phenotypes, which are influenced by genetic contexts. Thus, properly integrating genetic information and *APOE*-targeting interventions will enable early disease risk prediction for individuals and help stratify individuals for appropriate intervention strategies.

## Conflict of interest statement

Nothing declared.

## Acknowledgements

This work was supported in part by the Research Grants Council of Hong Kong (Theme-Based Research Scheme [T13-607/12R], and the Collaborative Research Fund [C6027-19GF]), the National Key R&D Program of China (2017YFE0190000 and 2018YFE0203600), the Areas of Excellence Scheme of the University Grants Committee (AoE/M-604/16), the Innovation and Technology Commission (ITCPD/17-9), the Guangdong Provincial Key S&T Program (2018B030336001), the Shenzhen Knowledge Innovation Program (JCYJ20180507183642005 and JCYJ20170413173717055), and the HKUST-SIAT Joint Laboratory for Brain Science.

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
  - of outstanding interest
- Shore VG, Shore B: **Heterogeneity of human plasma very low density lipoproteins. Separation of species differing in protein components.** *Biochemistry* 1973, **12**:502-507.
  - Shebline FA, Quarfordt SH: **A new apoprotein of human plasma very low density lipoproteins.** *J Biol Chem* 1974, **249**:1428-1433.
  - Utermann G: **Isolation and partial characterization of an arginine-rich apolipoprotein from human plasma very-low-density lipoproteins: apolipoprotein E.** *Hoppe Seylers Z Physiol Chem* 1975, **356**:1113-1122.
  - Utermann G, Menzel HJ, Langer KH: **On the polypeptide composition of an abnormal high density lipoprotein (LP-E) occurring in LCAT-deficient plasma.** *FEBS Lett* 1974, **45**:29-32.
  - Mahley RW: **Central nervous system lipoproteins: ApoE and regulation of cholesterol metabolism.** *Arterioscler Thromb Vasc Biol* 2016, **36**:1305-1315.
  - Getz GS, Reardon CA: **Apoprotein E as a lipid transport and signaling protein in the blood, liver, and artery wall.** *J Lipid Res* 2009, **50**:S156-S161.
  - Cooper AD: **Hepatic uptake of chylomicron remnants.** *J Lipid Res* 1997, **38**:2173-2192.
  - Huang Y, Mahley RW, Apolipoprotein E: **Structure and function in lipid metabolism, neurobiology, and Alzheimer's diseases.** *Neurobiol Dis* 2014, **72**:3-12.
  - Björkhem I, Meaney S, Fogelman AM: **Brain cholesterol: long secret life behind a barrier.** *Arterioscler Thromb Vasc Biol* 2004, **24**:806-815.
  - de Chaves EP, Narayanaswami V: **Apolipoprotein E and cholesterol in aging and disease in the brain.** *Future Lipidol* 2008, **3**:505-530.
  - Lo Sasso G, Schrage WK, Boué S, Veljkovic E, Peitsch MC, Hoeng J: **The Apoe-/- mouse model: a suitable model to study cardiovascular and respiratory diseases in the context of cigarette smoke exposure and harm reduction.** *J Transl Med* 2016, **14**:146.
  - Masliah E, Mallory M, Ge N, Alford M, Veinbergs I, Roses AD: **Neurodegeneration in the central nervous system of apoE-deficient mice.** *Exp Neurol* 1995, **136**:107-122.
  - Mak ACY, Pullinger CR, Tang LF, Wong JS, Deo RC, Schwarz JM, Gugliucci A, Movsesyan I, Ishida BY, Chu C et al.: **Effects of the absence of apolipoprotein E on lipoproteins, neurocognitive function, and retinal function.** *JAMA Neurol* 2014, **71**:1228-1236. By characterizing an individual harboring the homozygous *APOE* frame-shift mutation (c.291del, p.E97fs) who is devoid of *APOE* expression, the authors suggest that *APOE* is not essential for normal brain functioning. This work provides hints about the roles of *APOE* in the human system.
  - Schaum N, Karkanas J, Neff NF, May AP, Quake SR, Wyss-Coray T, Darmanis S, Batson J, Botvinnik O, Chen MB et al.: **Single-cell transcriptomics of 20 mouse organs creates a Tabula Muris.** *Nature* 2018, **562**:367-372.
  - Gaudreault N, Kumar N, Olivas VR, Eberlé D, Rapp JH, Raffai RL: **Macrophage-specific apoE gene repair reduces diet-induced hyperlipidemia and atherosclerosis in hypomorphic apoE mice.** *PLoS One* 2012, **7**.
  - Velmeshev D, Schirmer L, Jung D, Haeussler M, Perez Y, Mayer S, Bhaduri A, Goyal N, Rowitch DH, Kriegstein AR: **Single-cell genomics identifies cell type-specific molecular changes in autism.** *Science* (80-) 2019, **364**:685-689.
  - Casaletto KB, Elahi FM, Bettcher BM, Neuhaus J, Bendlin BB, Asthana S, Johnson SC, Yaffe K, Carlsson C, Blennow K et al.: **Neurogranin, a synaptic protein, is associated with memory independent of Alzheimer biomarkers.** *Neurology* 2017, **89**:1782-1788.
  - De Vos A, Jacobs D, Struyf H, Fransen E, Andersson K, Portelius E, Andreasson U, De Surgeloose D, Hernasteen D, Sleegers K et al.: **C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease.** *Alzheimers Dement* 2015, **11**:1461-1469.
  - Zhong L, Cherry T, Bies CE, Florence MA, Gerges NZ: **Neurogranin enhances synaptic strength through its interaction with calmodulin.** *EMBO J* 2009, **28**:3027-3039.
  - Zhang Y, Chen K, Sloan SA, Bennett ML, Scholze AR, O'Keeffe S, Phatnani HP, Guarnieri P, Caneda C, Ruderisch N et al.: **An RNA-seqencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex.** *J Neurosci* 2014, **34**:11929-11947.
  - Zhang Y, Sloan SA, Clarke LE, Caneda C, Plaza CA, Blumenthal PD, Vogel H, Steinberg GK, Edwards MSB, Li G et al.: **Purification and characterization of progenitor and mature human astrocytes reveals transcriptional and functional differences with mouse.** *Neuron* 2016, **89**:37-53.
  - Sun Y, Wu S, Bu G, Onifade MK, Patel SN, LaDu MJ, Fagan AM, Holtzman DM: **Glial fibrillary acidic protein-apolipoprotein E (apoE) transgenic mice: astrocyte-specific expression and differing biological effects of astrocyte-secreted apoE3 and apoE4 lipoproteins.** *J Neurosci* 1998, **18**:3261-3272.
  - Hoe HS, Harris DC, Rebeck GW: **Multiple pathways of apolipoprotein E signaling in primary neurons.** *J Neurochem* 2005, **93**:145-155.
  - Nathan BP, Jiang Y, Wong GK, Shen F, Brewer GJ, Struble RG: **Apolipoprotein E4 inhibits, and apolipoprotein E3 promotes neurite outgrowth in cultured adult mouse cortical neurons through the low-density lipoprotein receptor-related protein.** *Brain Res* 2002, **928**:96-105.
  - Huang YWA, Zhou B, Nabet AM, Wernig M, Südhof TC: **Differential signaling mediated by ApoE2, ApoE3, and ApoE4 in human neurons parallels Alzheimer's disease risk.** *J Neurosci* 2019, **39**:7408-7427.
  - Huang ZJ, Cao F, Wu Y, Peng JH, Zhong JJ, Jiang Y, Yin C, Guo ZD, Sun XC, Jiang L et al.: **Apolipoprotein E promotes white matter remodeling via the Dab1-dependent pathway after traumatic brain injury.** *CNS Neurosci Ther* 2020, **26**:698-710.

27. Veinbergs I, Everson A, Sagara Y, Masliah E: **Neurotoxic effects of apolipoprotein E4 are mediated via dysregulation of calcium homeostasis.** *J Neurosci Res* 2002, **67**:379-387.
28. Hayashi H, Campenot RB, Vance DE, Vance JE: **Protection of neurons from apoptosis by apolipoprotein e-containing lipoproteins does not require lipoprotein uptake and involves activation of phospholipase C $\gamma$ 1 and inhibition of calcineurin.** *J Biol Chem* 2009, **284**:29605-29613.
29. Srinivasan K, Friedman BA, Etxeberria A, Huntley MA, van der Brug MP, Foreman O, Paw JS, Modrusan Z, Beach TG, Serrano GE et al.: **Alzheimer's patient microglia exhibit enhanced aging and unique transcriptional activation.** *Cell Rep* 2020, **31**.
30. Krasemann S, Madore C, Cialic R, Baufeld C, Calcagno N, El Fatimy R, Beckers L, O'Loughlin E, Xu Y, Fanek Z et al.: **The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases.** *Immunity* 2017, **47**:566-581.e9
- By conducting transcriptomic analysis in microglia from human brains affected by Alzheimer's disease and microglia from multiple mouse models of diseases, the authors identified genes associated with neurodegenerative microglia phenotypes that are potentially modified by TREM2–APOE signaling. This work suggests that APOE plays roles in the transcriptional regulation of disease-associated gene expression in neurodegenerative diseases.
31. Theendakara V, Peters-Libeu CA, Spilman P, Poksay KS, Bredesen DE, Rao RV: **Direct transcriptional effects of apolipoprotein E.** *J Neurosci* 2016, **36**:685-700.
32. Theendakara V, Peters-Libeu CA, Bredesen DE, Rao RV: **Transcriptional effects of ApoE4: relevance to Alzheimer's disease.** *Mol Neurobiol* 2018, **55**:5243-5254.
33. Lee EG, Tulloch J, Chen S, Leong L, Saxton AD, Kraemer B, Darvas M, Keene CD, Shutes-David A, Todd K et al.: **Redefining transcriptional regulation of the APOE gene and its association with Alzheimer's disease.** *PLoS One* 2020, **15**
- By studying APOE transcripts in human postmortem brains, the authors identified the presence of a circular form of APOE transcript whose expression is altered in the brain in Alzheimer's disease. This is the first report of the presence of APOE circular RNA in the human system.
34. Sweeney MD, Sagare AP, Zlokovic BV: **Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders.** *Nat Rev Neurol* 2018, **14**:133-150.
35. Hafezi-Moghadam A, Thomas KL, Wagner DD: **ApoE deficiency leads to a progressive age-dependent blood-brain barrier leakage.** *Am J Physiol Physiol* 2007, **292**:C1256-C1262.
36. Bell RD, Winkler EA, Singh I, Sagare AP, Deane R, Wu Z, Holtzman DM, Betsholtz C, Armulik A, Sallstrom J et al.: **Apolipoprotein e controls cerebrovascular integrity via cyclophilin A.** *Nature* 2012, **485**:512-516.
37. Montagne A, Nation DA, Sagare AP, Barisano G, Sweeney MD, Chakhoyan A, Pachicano M, Joe E, Nelson AR, D'Orazio LM et al.: **APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline.** *Nature* 2020, **581**:71-76.
38. Blanchard JW, Bula M, Davila-Velderrain J, Akay LA, Zhu L, Frank A, Victor MB, Bonner JM, Mathys H, Lin YT et al.: **Reconstruction of the human blood-brain barrier in vitro reveals a pathogenic mechanism of APOE4 in pericytes.** *Nat Med* 2020, **26**:952-963
- By establishing an induced pluripotent stem cell-derived culture system mimicking the human blood–brain barrier, the authors investigated the mechanisms by which APOE-ε4 affects vascular function—specifically how it modulates amyloid accumulation via the blood–brain barrier. This study provides hints about how APOE-ε4 affects the blood–brain barrier in neurodegenerative diseases.
39. Kraft HG, Menzel HJ, Hoppichler F, Vogel W, Utermann G: **Changes of genetic apolipoprotein phenotypes caused by liver transplantation. Implications for apolipoprotein synthesis.** *J Clin Invest* 1989, **83**:137-142.
40. Linton MF, Gish R, Hubl ST, Büttler E, Esquivel C, Bry WI, Boyles JK, Wardell MR, Young SG: **Phenotypes of apolipoprotein B and apolipoprotein E after liver transplantation.** *J Clin Invest* 1991, **88**:270-281.
41. Zlokovic BV, Martel CL, Mackie JB, Matsubara E, Wisniewski T, McComb JG, Frangione B, Ghiso J: **Brain uptake of circulating apolipoproteins J and E complexed to Alzheimer's amyloid β.** *Biochem Biophys Res Commun* 1994, **205**:1431-1437.
42. Lane-Donovan C, Wong WM, Durakoglugil MS, Wasser CR, Jiang S, Xian X, Herz J: **Genetic restoration of plasma apoE improves cognition and partially restores synaptic defects in ApoE-deficient mice.** *J Neurosci* 2016, **36**:10141-10150
- By comparing the cognition and synaptic activity between ApoE-general knockout mice and ApoE brain-specific—knockout mice, the authors observed that restoring plasma ApoE improves learning, memory, and synaptic function. This work suggests that peripheral APOE plays a role in modifying brain cognition and neuronal functions.
43. Zannis VI, Breslow JL: **Human very low density lipoprotein apolipoprotein E isoprotein polymorphism is explained by genetic variation and posttranslational modification.** *Biochemistry* 1981, **20**:1033-1041.
44. Zannis VI, Breslow JL, Utermann G, Mahley RW, Weisgraber KH, Havel RJ, Goldstein JL, Brown MS, Schonfeld G, Hazzard WR et al.: **Proposed nomenclature of apoE isoproteins, apoE genotypes, and phenotypes.** *J Lipid Res* 1982, **23**:911-914.
45. Utermann G, Steinmetz A, Weber W: **Genetic control of human apolipoprotein E polymorphism: comparison of one-and two-dimensional techniques of isoprotein analysis.** *Hum Genet* 1982, **60**:344-351.
46. Weisgraber KH, Rall SC, Mahley RW: **Human E apoprotein heterogeneity. Cysteine-arginine interchanges in the amino acid sequence of the apo-E isoforms.** *J Biol Chem* 1981, **256**:9077-9083.
47. Hatters DM, Peters-Libeu CA, Weisgraber KH: **Apolipoprotein E structure: insights into function.** *Trends Biochem Sci* 2006, **31**:445-454.
48. Fernandez CG, Hamby ME, McReynolds ML, Ray WJ: **The role of apoE4 in disrupting the homeostatic functions of astrocytes and microglia in aging and Alzheimer's disease.** *Front Aging Neurosci* 2019, **10**.
49. Dong LM, Wilson C, Wardell MR, Simmons T, Mahley RW, Weisgraber KH, Agard DA: **Human apolipoprotein E. Role of arginine 61 in mediating the lipoprotein preferences of the E3 and E4 isoforms.** *J Biol Chem* 1994, **269**:22358-22365.
50. Mahley RW, Rall SC, Apolipoprotein E: **Far more than a lipid transport protein.** *Annu Rev Genomics Hum Genet* 2000, **1**:507-537.
51. Dong LM, Weisgraber KH: **Human apolipoprotein E4 domain interaction. Arginine 61 and glutamic acid 255 interact to direct the preference for very low density lipoproteins.** *J Biol Chem* 1996, **271**:19053-19057.
52. Utermann G, Hees M, Steinmetz A: **Polymorphism of apolipoprotein E and occurrence of dysbetalipoproteinemia in man.** *Nature* 1977, **269**:604-607.
53. Suri S, Heise V, Trachtenberg AJ, Mackay CE: **The forgotten APOE allele: a review of the evidence and suggested mechanisms for the protective effect of APOE ε2.** *Neurosci Biobehav Rev* 2013, **37**:2878-2886.
54. Verghese PB, Castellano JM, Holtzman DM: **Apolipoprotein E in Alzheimer's disease and other neurological disorders.** *Lancet Neurol* 2011, **10**:241-252.
55. Liu CC, Kanekiyo T, Xu H, Bu G: **Apolipoprotein e and Alzheimer disease: risk, mechanisms and therapy.** *Nat Rev Neurol* 2013, **9**:106-118.
56. Kim J, Yoon H, Basak J, Kim J: **Apolipoprotein E in synaptic plasticity and alzheimer's disease: potential cellular and molecular mechanisms.** *Mol Cells* 2014, **37**:833-840.
57. Najm R, Jones EA, Huang Y: **Apolipoprotein E4, inhibitory network dysfunction, and Alzheimer's disease.** *Mol Neurodegener* 2019, **14**.

58. Piers R: Structural brain volume differences between cognitively intact ApoE4 carriers and non-carriers across the lifespan. *Neural Regen Res* 2018, **13**:1309-1312.
59. Rawle MJ, Davis D, Bendayan R, Wong A, Kuh D, Richards M: **Apolipoprotein-E (ApoE) ε4 and cognitive decline over the adult life course.** *Transl Psychiatry* 2018, **8**.
60. Blacker D, Haines JL, Rodes L, Terwedow H, Go RCP, Harrell LE, Perry RT, Bassett SS, Chase G, Meyers D et al.: **ApoE-4 and age at onset of Alzheimer's disease: the NIMH genetics initiative.** *Neurology* 1997, **48**:139-147.
61. Keene CD, Cudaback E, Li X, Montine KS, Montine TJ: **Apolipoprotein E isoforms and regulation of the innate immune response in brain of patients with Alzheimer's disease.** *Curr Opin Neurobiol* 2011, **21**:920-928.
62. Kanekiyo T, Xu H, Bu G: **ApoE and Aβ in Alzheimer's disease: accidental encounters or partners?** *Neuron* 2014, **81**:740-754.
63. Shi Y, Yamada K, Liddelow SA, Smith ST, Zhao L, Luo W, Tsai RM, Spina S, Grinberg LT, Rojas JC et al.: **ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy.** *Nature* 2017, **549**:523-527.
64. Iron A, Richard P, Pascual de Zulueta M, Thomas G, Thomas M: **Genotyping of a patient homozygous for a rare apolipoprotein E1 [Gly127→Asp; Arg158→Cys] (Weisgraber allele).** *J Inher Metab Dis* 1995, **18**:723-726.
65. Murrell JR, Price BM, Baiyewu O, Gureje O, Deeg M, Hendrie H, Ogunniyi A, Hall K: **The fourth apolipoprotein E haplotype found in the Yoruba of Ibadan.** *Am J Med Genet Part B Neuropsychiatr Genet* 2006, **141**:426-427.
66. Maeda H, Nakamura H, Kobori S, Okada M, Niki H, Ogura T, Hiraga S: **Molecular cloning of a human apolipoprotein E variant: E5 (Glu3—Lys3).** *J Biochem* 1989, **105**:491-493.
67. Wardell MR, Rall SC, Schaefer EJ, Kane JP, Weisgraber KH: **Two apolipoprotein E5 variants illustrate the importance of the position of additional positive charge on receptor-binding activity.** *J Lipid Res* 1991, **32**:521-528.
68. Maeda H, Nakamura H, Kobori S, Okada M, Mori H, Niki H, Ogura T, Hiraga S: **Identification of human apolipoprotein E variant gene: apolipoprotein E7 (Glu244,245↑Lys244,245).** *J Biochem* 1989, **105**:51-54.
69. Wardell MR, Brennan SO, Janus ED, Fraser R, Carrell RW: **Apolipoprotein E2-Christchurch (136 Arg—Ser). New variant of human apolipoprotein E in a patient with type III hyperlipoproteinemia.** *J Clin Invest* 1987, **80**:483-490.
70. Georgiadou D, Chroni A, Vezeridis A, Zannis VI, Stratikos E: **Biophysical analysis of apolipoprotein E3 variants linked with development of type III hyperlipoproteinemia.** *PLoS One* 2011, **6**.
71. Lazar A, Weisgraber KH, Rall SC, Giladi H, Innerarity TL, Levanon AZ, Boyles JK, Amit B, Gorecki M, Mahley RW et al.: **Site-specific mutagenesis of human apolipoprotein E. Receptor binding activity of variants with single amino acid substitutions.** *J Biol Chem* 1988, **263**:3542-3545.
72. Arboleda-Velasquez JF, Lopera F, O'Hare M, Delgado-Tirado S, Marino C, Chmielewska N, Saez-Torres KL, Amarnani D, Schultz AP, Sperling RA et al.: **Resistance to autosomal dominant Alzheimer's disease in an APOE3 Christchurch homozygote: a case report.** *Nat Med* 2019, **25**:1680-1683.
- The authors characterized an individual harboring the pathogenic *PSEN1* E280A mutation and 2 copies of the *APOE*-ε3 Christchurch (R136S) mutation. The individual exhibited a high amyloid burden accompanied by mild tau deposition and neurodegeneration in the brain. This work suggests that *APOE* plays roles in disease pathogenesis and progression, and is a potential target for disease intervention.
73. Foraker J, Millard SP, Leong L, Thomson Z, Chen S, Keene CD, Bekris LM, Yu CE, Fischer A: **The APOE gene is differentially methylated in Alzheimer's disease.** *J Alzheimer's Dis* 2015, **48**:745-755.
74. Yu CE, Cudaback E, Foraker J, Thomson Z, Leong L, Lutz F, Gill JA, Saxton A, Kraemer B, Navas P et al.: **Epigenetic signature and enhancer activity of the human APOE gene.** *Hum Mol Genet* 2013, **22**:5036-5047.
75. Bertrand P, Poirier J, Oda T, Finch CE, Pasinetti GM: **Association of apolipoprotein E genotype with brain levels of apolipoprotein E and apolipoprotein J (clusterin) in Alzheimer disease.** *Mol Brain Res* 1995, **33**:174-178.
76. Zhou X, Chen Y, Mok KY, Kwok TCY, Mok VCT, Guo Q, Ip FC, Chen Y, Mullapudi N, Weiner MW et al.: **Non-coding variability at the APOE locus contributes to the Alzheimer's risk.** *Nat Commun* 2019, **10**
- By conducting fine-mapping analysis using whole-genome sequencing data of *APOE* and nearby regions, the authors identified noncoding variants and haplotypes that exert disease risk effects independent of *APOE*-ε4. Genotype-expression and genotype-endophenotype association analyses further suggest the roles of those noncoding risk variants in modifying *APOE* expression and Alzheimer's disease-associated endophenotypes. This work resolves the debate on the roles of noncoding risk variants in *APOE* and nearby regions, and suggests an alternative disease mechanism in parallel with *APOE*-ε4 coding risk.
77. Lambert JC, Pérez-Tur J, Dupire MJ, Galasko D, Mann D, Amouyel P, Hardy J, Delacourte A, Chartier-Harlin MC: **Distortion of allelic expression of apolipoprotein E in Alzheimer's disease.** *Hum Mol Genet* 1997, **6**:2151-2154.
78. Najm R, Jones EA, Huang Y: **Apolipoprotein E4, inhibitory network dysfunction, and Alzheimer's disease.** *Mol Neurodegener* 2019, **14**:24.
79. Bekris LM, Millard SP, Galloway NM, Vuletic S, Albers JJ, Li G, Galasko DR, DeCarli C, Farlow MR, Clark CM et al.: **Multiple SNPs within and surrounding the apolipoprotein E gene influence cerebrospinal fluid apolipoprotein E protein levels.** *J Alzheimer's Dis* 2008, **13**:255-266.
80. Bekris LM, Galloway NM, Montine TJ, Schellenberg GD, Yu CE: ***APOE* mRNA and protein expression in postmortem brain are modulated by an extended haplotype structure.** *Am J Med Genet Part B Neuropsychiatr Genet* 2010, **153**:409-417.
81. Laws SM, Hone E, Gandy S, Martins RN: **Expanding the association between the *APOE* gene and the risk of Alzheimer's disease: possible roles for *APOE* promoter polymorphisms and alterations in *APOE* transcription.** *J Neurochem* 2003, **84**:1215-1236.
82. Bullido MJ, Artiga MJ, Recuero M, Sastre I, García MA, Aldudo J, Lendon C, Han SW, Morris JC, Frank A et al.: **A polymorphism in the regulatory region of *APOE* associated with risk for Alzheimer's dementia.** *Nat Genet* 1998, **18**:69-71.
83. Bratosiewicz-Wasik J, Liberski PP, Peplonska B, Styczynska M, Smolen-Dzirba J, Cycon M, Wasik TJ: **Regulatory region single nucleotide polymorphisms of the apolipoprotein E gene as risk factors for Alzheimer's disease.** *Neurosci Lett* 2018, **684**:86-90.
84. Zhou X, Chen Y, Mok KY, Zhao Q, Chen K, Chen Y, Hardy J, Li Y, Fu AKY, Guo Q et al.: **Identification of genetic risk factors in the Chinese population implicates a role of immune system in Alzheimer's disease pathogenesis.** *Proc Natl Acad Sci U S A* 2018, **115**:1697-1706.
85. Jansen IE, Savage JE, Watanabe K, Bryois J, Williams DM, Steinberg S, Sealock J, Karlsson IK, Hägg S, Athanasiu L et al.: **Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk.** *Nat Genet* 2019, **51**:404-413.
86. Takei N, Miyashita A, Tsukie T, Arai H, Asada T, Imagawa M, Shoji M, Higuchi S, Urakami K, Kimura H et al.: **Genetic association study on in and around the *APOE* in late-onset Alzheimer disease in Japanese.** *Genomics* 2009, **93**:441-448.
87. Babenko VN, Afonnikov DA, Ignatieva EV, Klimov AV, Gusev FE, Rogaev EI: **Haplotype analysis of *APOE* intragenic SNPs.** *BMC Neurosci* 2018, **19**
- By conducting genetic and haplotype analyses in different ethnic groups, the authors identified Alzheimer's disease-associated haplotypes whose prevalence varies among ethnic groups. This work suggests that ethnic background might modify the risk effects of *APOE* by influencing the genomic content of *APOE* and nearby genomic regions.

88. Rajabli F, Feliciano BE, Celis K, Hamilton-Nelson KL, Whitehead PL, Adams LD, Bussies PL, Manrique CP, Rodriguez A, Rodriguez V et al.: **Ancestral origin of ApoE ε4 Alzheimer disease risk in Puerto Rican and African American populations.** *PLoS Genet* 2018, **14**
- By conducting genetic analyses with 2 different ethnic groups, the authors revealed the roles of regions near *APOE* in modulating Alzheimer's disease risk associated with *APOE*-ε4. The study presents an explanation for the observed variations in Alzheimer's disease risk associated with *APOE*-ε4 across populations.
89. Tang MX, Stern Y, Marder K, Bell K, Gurland B, Lantigua R, Andrews H, Feng L, Tycko B, Mayeux R: **The APOE-ε4 allele and the risk of Alzheimer disease among African Americans, whites, and Hispanics.** *J Am Med Assoc* 1998, **279**:751-755.
90. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM et al.: **Neuroinflammation in Alzheimer's disease.** *Lancet Neurol* 2015, **14**:388-405.
91. Sochocka M, Zwolińska K, Leszek J: **The infectious etiology of Alzheimer's disease.** *Curr Neuropharmacol* 2017, **15**:996-1009.
92. Yoo S-M, Park J, Kim S-H, Jung Y-K: **Emerging perspectives on mitochondrial dysfunction and inflammation in Alzheimer's disease.** *BMB Rep* 2020, **53**:35-46.
93. Balin BJ, Hudson AP: **Herpes viruses and Alzheimer's disease: new evidence in the debate.** *Lancet Neurol* 2018, **17**:839-841.
94. Cairns DM, Rouleau N, Parker RN, Walsh KG, Gehrkne L, Kaplan DL: **A 3D human brain-like tissue model of herpes-induced Alzheimer's disease.** *Sci Adv* 2020, **6**.
95. Wang W, Zhao F, Ma X, Perry G, Zhu X: **Mitochondria dysfunction in the pathogenesis of Alzheimer's disease: recent advances.** *Mol Neurodegener* 2020, **15**.
96. Fuior EV, Gafencu AV: **Apolipoprotein c1: its pleiotropic effects in lipid metabolism and beyond.** *Int J Mol Sci* 2019, **20**.
97. Yin C, Ackermann S, Ma Z, Mohanta SK, Zhang C, Li Y, Nietzsche S, Westermann M, Peng L, Hu D et al.: **ApoE attenuates unresolvable inflammation by complex formation with activated C1q.** *Nat Med* 2019, **25**:496-506.
98. Cramer PE, Cirrito JR, Wesson DW, Lee CYD, Karlo JC, Zinn AE, Casali BT, Restivo JL, Goebel WD, James MJ et al.: **ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models.** *Science* (80-) 2012, **335**:1503-1506.
99. Wang C, Najm R, Xu Q, Jeong DE, Walker D, Balestra ME, Yoon SY, Yuan H, Li G, Miller ZA et al.: **Gain of toxic apolipoprotein E4 effects in human iPSC-derived neurons is**

**ameliorated by a small-molecule structure corrector article.**  
*Nat Med* 2018, **24**:647-657

By studying induced pluripotent stem cell-derived human neurons harboring the *APOE*-ε3 or *APOE*-ε4 genotype, the authors revealed how *APOE*-ε4 modifies neuronal functions. Specifically, the authors demonstrate the beneficial effects of an ApoE4-targeting small-molecule structure corrector, which alleviates the phenotypes in neurons harboring *APOE*-ε4. Thus, this study suggests a possible intervention strategy that involves targeting ApoE proteins.

100. Liao F, Li A, Xiong M, Bien-Ly N, Jiang H, Zhang Y, Finn MB, Hoyle R, Keyser J, Lefton KB et al.: **Targeting of nonlipidated, aggregated apoE with antibodies inhibits amyloid accumulation.** *J Clin Invest* 2018, **128**:2144-2155.
101. Huynh TPV, Liao F, Francis CM, Robinson GO, Serrano JR, Jiang H, Roh J, Finn MB, Sullivan PM, Esparza TJ et al.: **Age-dependent effects of apoE reduction using antisense oligonucleotides in a model of β-amyloidosis.** *Neuron* 2017, **96**:1013-1023.e4.
102. Hu J, Liu CC, Chen XF, Zhang YW, Xu H, Bu G: **Opposing effects of viral mediated brain expression of apolipoprotein E2 (apoE2) and apoE4 on apoE lipidation and Aβ metabolism in apoE4-targeted replacement mice.** *Mol Neurodegener* 2015, **10**.
103. Cummings JL, Zhong K, Kinney JW, Heaney C, Moll-Tudla J, Joshi A, Pontecorvo M, Devous M, Tang A, Bena J: **Double-blind, placebo-controlled, proof-of-concept trial of bexarotene Xin moderate Alzheimer's disease.** *Alzheimers Res Ther* 2016, **8**:4
- The authors examined the brain amyloid load in patients receiving the retinoid X receptor agonist bexarotene and observed variable effects of bexarotene on brain amyloid load between *APOE*-ε4 carriers and non-carriers. Specifically, they did not observe changes in amyloid burden among *APOE*-ε4 carriers after bexarotene treatment. The study suggests that certain genotypes modify drug responses.
104. Luo W, Schork NJ, Marschke KB, Ng SC, Hermann TW, Zhang J, Sanders JM, Tooker P, Malo N, Zapala MA et al.: **Identification of polymorphisms associated with hypertriglyceridemia and prolonged survival induced by bexarotene in treating non-small cell lung cancer.** *Anticancer Res* 2011, **31**:2303-2311.
105. Escott-Price V, Myers A, Huentelman M, Shoai M, Hardy J: **Polygenic risk score analysis of Alzheimer's disease in cases without APOE4 or APOE2 alleles.** *J Prev Alzheimers Dis* 2019, **6**:16-19.
106. Escott-Price V, Shoai M, Pither R, Williams J, Hardy J: **Polygenic score prediction captures nearly all common genetic risk for Alzheimer's disease.** *Neurobiol Aging* 2017, **49**:214.e7.
107. Zhou X, Chen Y, Ip FCF, Lai NCH, Li YYT, Jiang Y, Zhong H, Chen Y, Zhang Y, Ma S et al.: **Genetic and polygenic risk score analysis for Alzheimer's disease in the Chinese population.** *Alzheimers Dement* 2020, **12**:e12074.